



DREXEL UNIVERSITY

College of
Medicine

Gene Expression Analysis of Atypical Fibroxanthoma Patients Reveals New Insights into the Tumor Microenvironment

Nia Gyongyosi, B.S.* , Aya Al-Nazal, B.S., M.S.* , and Shehbeel Arif, B.S., M.S.

Drexel University College of Medicine, Philadelphia, PA, USA



TOWER HEALTH

Advancing Health. Transforming Lives.

No conflict of interest declared.
No IRB/IACUC review needed.

Introduction

Atypical Fibroxanthoma

- Rare low grade superficial carcinoma

Background

- Less aggressive than undifferentiated pleomorphic sarcoma
- Sun exposed areas of the body
- Ultraviolet light induction
- Mohs surgery

Cell origin

- Unclear
- Previously proposed Fibroblast origin

Purpose

- Expand current knowledge on AFX microenvironment and tumor origin
- Analysis of data set comprising 8 superficial cutaneous papules and nodules of AFX excised by Mohs surgery and subjected to cell deconvolution analysis and pathway analysis.²

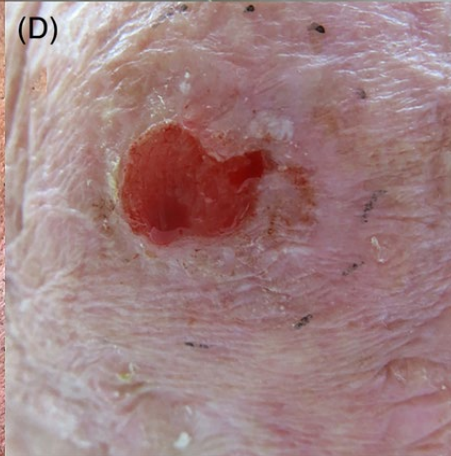
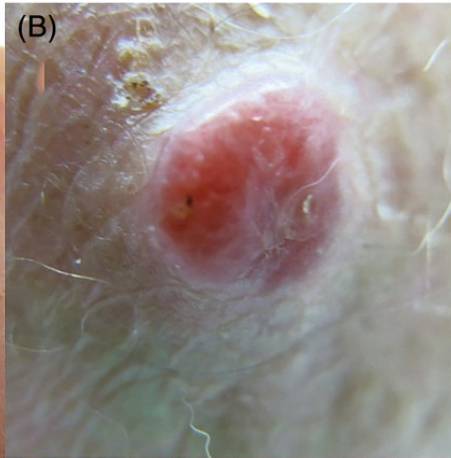
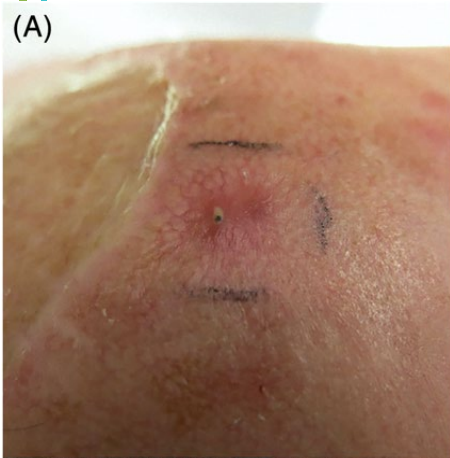
Hypothesis

- We hypothesized that the cellular origin of AFX is of epithelial or keratinocyte origin.



2. Lai, K., et al. (2017). Genomic analysis of atypical fibroxanthoma. *PLoS one*, 12(11), e0188272.

Clinical Presentation



(A) Ill-defined small plaque on the scalp.

(B) Flesh-colored nodule surrounded by actinic keratoses.

(C) Keratotic nodule on the tragus.

(D) Eroded plaque on elastotic scalp.

Methods

RNA-Sequencing paired expression data on AFX by Lai *et al.* (GSE85671)

- We analyzed publicly available RNA-sequencing data from eight AFX patient tumor tissues and paired histologically normal tissues bordering the lesion after successful excision by Mohs surgery.

Cell Deconvolution

- Analyze genetic component of cells to ascertain cell origin
- Analysis based on RNA sequencing
- Analysis was performed using xCell package in R to ascertain AFX cell-type composition and immune cell microenvironment as well as controls.
- Gene analysis utilized for computational estimation of different cell types.

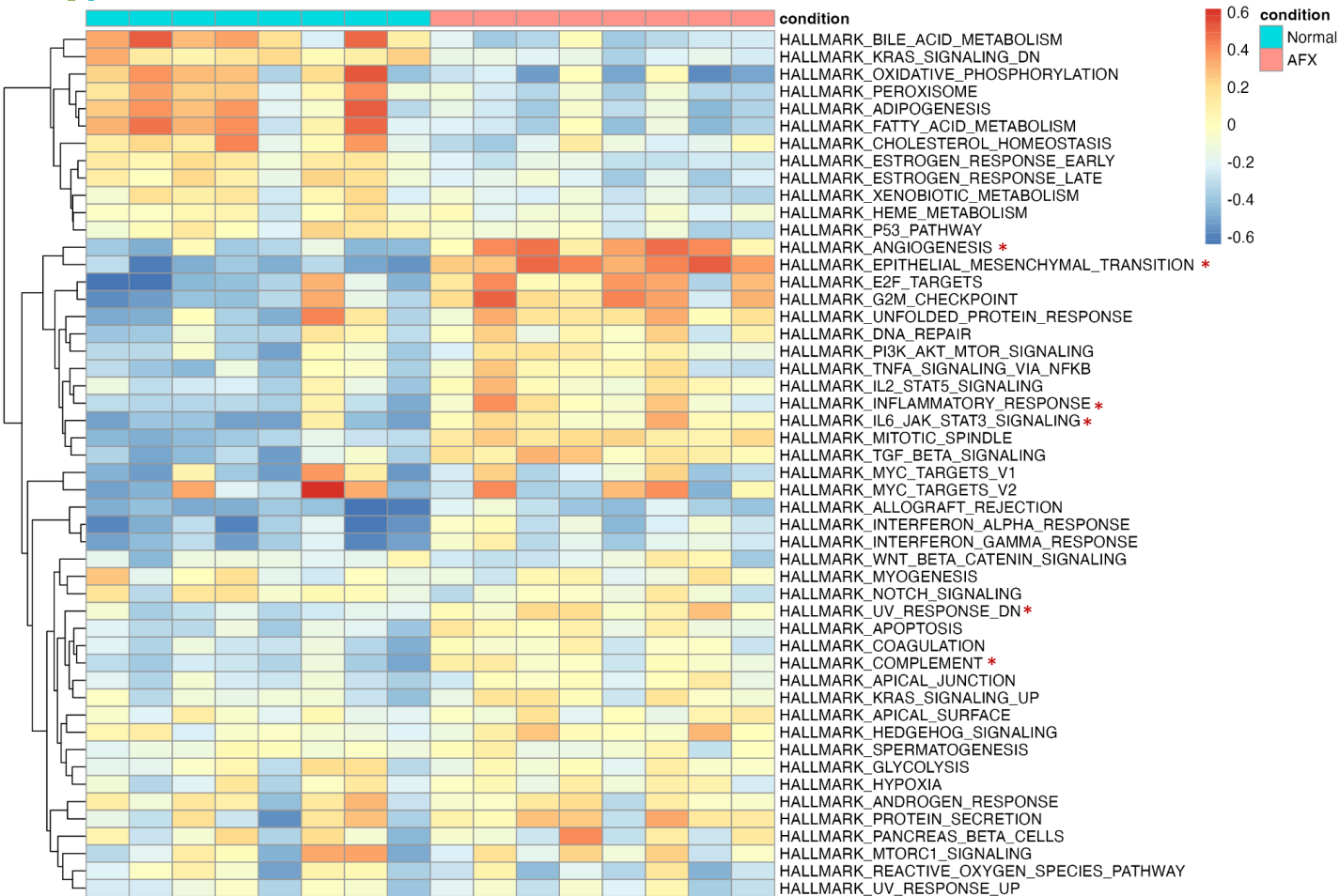
Pathway Analysis (GSVA)

- Used to analyze the cell makeup of the tumor tissue compared to normal tissue
- Gene set variation analysis (GSVA) utilized to pinpoint cellular pathways upregulated or downregulated
- Whether gene set is upregulated or downregulated as a whole

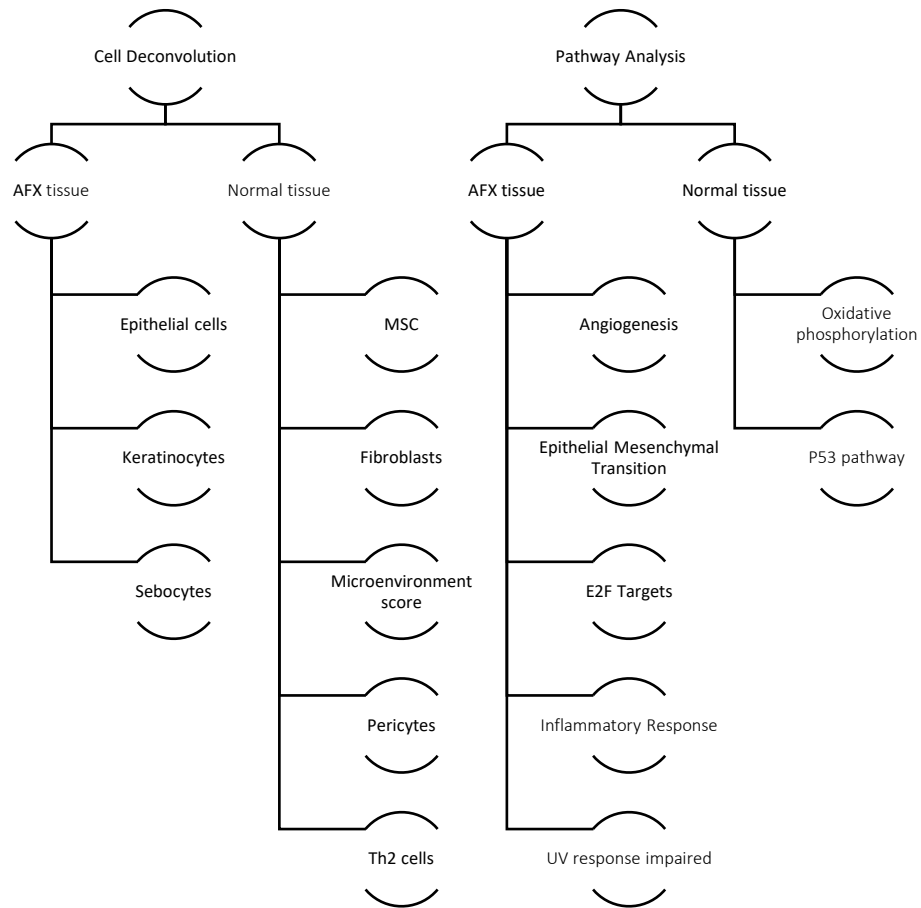
Cell Deconvolution



Pathway Analysis (GSVA)



Results



Discussion

- The cellular origin of AFX is currently unclear. The characteristic spindled cells found in AFX tissue have been hypothesized to be EMT-transitioned keratinocytes (Mirza & Weedon, 2005; Nakamura *et al.*, 2010)
- Our cell deconvolution revealed an enriched presence of Epithelial cells, keratinocytes, and sebocytes within the AFX tissue lesions.
- On the contrary, the bordering normal tissue had enrichment of Mesenchymal Stem Cells, and Fibroblasts, as well as Type II Helper T cells.
- EMT and Inflammatory response pathways were upregulated in AFX tissue.
- There was no infiltration of macrophages or other immune cells revealed by cell deconvolution, which was otherwise reported previously by the Arron group (Lai *et al.*, 2017).

Conclusion and Future Prospect

- Our computational analyses suggest that AFX may be of epithelial rather than fibroblast origin.
- Presence of immune cell infiltration surrounding AFX tumors.
- Increase in the presence of inflammatory pathways in AFX tumor tissue.
- Expanding on previous findings is integral to develop treatment approaches and to better understand the prognosis of AFX.
- Future prospects would focus on investigating mitochondrial dysregulation and the possible molecular drivers of AFX.

References

- Ak, M., Kahraman, A., Arnold, F. M., Turko, P., Levesque, M. P., Zoche, M., Ramelyte, E., & Dummer, R. (2021). Clinicopathological and Genomic Profiles of Atypical Fibroxanthoma and Pleomorphic Dermal Sarcoma Identify Overlapping Signatures with a High Mutational Burden. *Genes*, 12(7), 974. <https://doi.org/10.3390/genes12070974>
- Bitel, Alena & Schönlebe, Jacqueline & Krönert, Claudia & Wollina, Uwe. (2020). Atypical Fibroxanthoma – An Analysis of 105 Tumors. *Dermatologic Therapy*. 33. 10.1111/dth.13962.
- Aran, D., Hu, Z., & Butte, A. J. (2017). xCell: digitally portraying the tissue cellular heterogeneity landscape. *Genome biology*, 18(1), 220. <https://doi.org/10.1186/s13059-017-1349-1>
- Hänzelmann, S., Castelo, R., & Guinney, J. (2013). GSEA: gene set variation analysis for microarray and RNA-seq data. *BMC bioinformatics*, 14, 7. <https://doi.org/10.1186/1471-2105-14-7>
- Lai, K., Harwood, C. A., Purdie, K. J., Proby, C. M., Leigh, I. M., Ravi, N., Mully, T. W., Brooks, L., Sandoval, P. M., Rosenblum, M. D., & Arron, S. T. (2017). Genomic analysis of atypical fibroxanthoma. *PloS one*, 12(11), e0188272. <https://doi.org/10.1371/journal.pone.0188272>
- Mirza, B., & Weedon, D. (2005). Atypical fibroxanthoma: a clinicopathological study of 89 cases. *The Australasian journal of dermatology*, 46(4), 235–238. <https://doi.org/10.1111/j.1440-0960.2005.00190.x>
- Nakamura, M., Sugita, K., & Tokura, Y. (2010). Expression of Snail1 in the vimentin-expressing squamous cell carcinoma mimicking atypical fibroxanthoma: possible involvement of an epithelial-mesenchymal transition. *Journal of the European Academy of Dermatology and Venereology : JEADV*, 24(11), 1365–1366. <https://doi.org/10.1111/j.1468-3083.2010.03659.x>
- Pfeifer G. P. (2020). Mechanisms of UV-induced mutations and skin cancer. *Genome instability & disease*, 1(3), 99–113. <https://doi.org/10.1007/s42764-020-00009-8>